

**COMPOSITIONS AND METHODS FOR DETECTING AND QUANTIFYING  
COX-2 ACTIVITY AND ANANDAMIDE METABOLITES**

**APPLICATION FOR UNITED STATES LETTERS PATENT**

This application claims benefit of U.S. Patent Application Serial No. 60/223,665 filed August 7, 2000, entitled "Method for *in vitro* and *in vivo* determination of COX-2 activity" and U.S. Patent Application Serial No. 60/302,975 filed July 03, 2001, entitled "COX-2 Assays"; each application incorporated herein by reference.

Be it known that we, Lawrence J. Marnett, a citizen of the United States, residing at 1884 Laurel Ridge Drive, Nashville, TN 37215, and Kevin R. Kozak, a citizen of the United States, residing at 1906 Chet Atkins Blvd. #705, Nashville, TN 37212; have invented a new and useful "Compositions and Methods for Detecting and Quantifying COX-2 Activity and Anandamide Metabolites."

**GOVERNMENT SUPPORT CLAUSE**

This invention was made with federal grant 5R35CA47479, and with partial support from the National Foundation for Cancer Research. The United States Government has certain rights in this invention.

## TABLE OF CONTENTS

	1.	Background of the Invention	
	1.1	Field of the Invention	
	1.2	Description of the Relevant Art	
5	2.	Summary of the Invention	
	3.	Brief Description of the Drawings	
	4.	Detailed Description of the Invention	
	4.1	Definitions	
	4.2	Cyclooxygenases and Prostaglandins	
10	4.3	Detecting COX-2 Activity by Detecting Prostaglandin Ethanolamides	
	4.4	Measuring Prostaglandin Ethanolamides	
	4.4.1	Use of a Standard	
	4.4.2	Detection and Measuring Devices	
	4.4.3	Separation Devices	
15	4.4.4	Subjects	
	4.4.5	Samples	
	4.4.6	Antibodies	
	4.5	COX-2 and Inflammatory Diseases	
	4.6	COX-2 and Cancer	
20	4.7	COX-2 and Research	
	4.8	Kits	
	5.0	Examples	
	5.1	Example 1	
	5.2	Example 2	
25	5.3	Example 3	
	5.4	Example 4	
	5.5	Example 5	

5.6 Example 6

5.7 Example 7

## 1. BACKGROUND OF THE INVENTION

### 5 1.1 FIELD OF THE INVENTION

The present invention relates generally to the cyclooxygenases and their roles in human pathology, including cancer and inflammation. More particularly, this invention pertains to methods and articles of manufacture for detecting or measuring COX-2 activity by detecting and measuring COX-2 specific enzymatic products including prostaglandin ethanolamides and thromboxane ethanolamides.

### 1.2 DESCRIPTION OF THE RELATED ART

COX is a prostaglandin endoperoxide synthase enzyme (cyclooxygenase, COX, EC 1.14.99.1), which catalyzes the conversion of arachidonic acid to prostaglandin (PG) H<sub>2</sub>. Two isoforms of COX are known, COX-1 and COX-2. COX-1 is constitutively expressed. COX-2, however, is inducible in a variety of cells, especially those of the central nervous and immune systems (Masferrer *et al.* 1994, *Proc. Natl. Acad. Sci. USA* 91:3228-3232; Vane *et al.* 1994, *Proc. Natl. Acad. Sci. USA* 91:2046-2050; Kennedy *et al.* 1993, *Biochem. Biophys. Res. Commun.* 197:494-500). Certain changes in COX-2 activity are associated with a variety of human inflammatory diseases. These diseases include, but are not limited to, acute appendicitis, asthma, myocardial infarction, certain immunological disease processes, infection, malignancy, endotoxemia and reperfusion injury. In addition, inappropriate COX-2 expression or over-expression is associated with certain types of cancers, including, but not limited to, carcinoma of the colon, rectum, stomach, esophagus, lung, and skin. The amount of COX-2 expression is related to the stage or progression of cancer (Fosslien, E, *et al.* 2000, *Ann. Clin. Lab. Sci.* 30:3-21). COX-

2 has become a major pharmaceutical target for developing treatments for these and other diseases. Methods of detecting and measuring COX-2 activity are highly desired.

5 Yu *et al.* (1997) J. Biol. Chem. 272:21181-21186, describes the enzymatic conversion of arachidonyl ethanolamide (anandamide, AEA), to PGE<sub>2</sub>-ethanolamide in cell lines expressing COX-2 but not COX-1.

10 U.S. Patent 5,543,297 to Cromlish *et al.*, describes measuring total COX activity (COX-1 activity and COX-2 activity) in separate samples, with and without a COX-2 specific inhibitor, and then indirectly estimating COX-2 specific activity by subtracting the total COX activity observed with the inhibitor from the total COX activity observed without the inhibitor. One major weakness of this method is that the dynamics of enzymatic inhibition change based upon numerous variables including time, temperature, concentration, specificity, sample preparation, etc.

15 U.S. patent 5,475,021 to Marnett *et al.* describes a method of measuring the activity of purified COX-2 by measuring O<sub>2</sub>-uptake during catalysis. This method requires purification of the enzyme.

20 U.S. Patent 6,045,773 to Isakson *et al.*, describes a method for measuring COX-2 activity in a mammal by administering a positron-emitting radioisotope-labeled COX-2 selective-binding agent to the mammal and then detecting the label by positron-emission tomography (PET). Weaknesses of this method include the invasive nature and expense of PET equipment. In addition, the method only localizes COX-2 protein but does not detect or measure activity.

25 What is needed, then, is a less-invasive, direct method of selectively detecting and measuring COX-2 activity in biological samples and whole animals without the need to purify the enzyme.

## 2. SUMMARY OF THE INVENTION

The present invention provides, in part, articles of manufacture and methods for selectively detecting and measuring COX-2 enzymatic activity. One object of the invention is to provide a system for distinguishing between COX-2 activity and  
5 COX-1 activity. COX-1 and COX-2 are well known to catalyze the committed step in the conversion of arachidonic acid to PGH<sub>2</sub>. Downstream events, both enzymatic and non-enzymatic, further convert the PGH<sub>2</sub> into a variety of prostaglandins and thromboxanes. The present invention provides herein a system and methods to exploit a COX-2 selective enzymatic reaction with AEA for specifically detecting and  
10 measuring COX-2 enzymatic activity. In certain embodiments of the present invention, COX-2 enzymatic activity is detected or measured by detecting or measuring novel PGH<sub>2</sub>-EA metabolites, including, but not limited to: PGB<sub>2</sub>-EA, PGD<sub>2</sub>-EA, PGE<sub>2</sub>-EA, PGF<sub>2</sub> $\alpha$ -EA, TxB<sub>2</sub>-EA, 6-keto-PGF<sub>1</sub> $\alpha$ -EA, 15-keto-PGE<sub>2</sub>-EA, 13,14-dihydro-15-keto-PGE<sub>2</sub>-EA, PGG<sub>2</sub>-EA, PGH<sub>2</sub>-EA, PGA<sub>2</sub>-EA, PGJ<sub>2</sub>-EA, PGJ<sub>2</sub>-EA  
15 derivatives, bicyclo-PGE<sub>2</sub>-EA, TxA<sub>2</sub>-EA and PGI<sub>2</sub>-EA. In certain embodiments, the COX-2 specific enzymatic activity is compared to COX-1 activity, for example, a COX-2/COX-1 ratio is reported.

One aspect of the present invention is a method of detecting COX-2 activity in a biological system or subject comprising detecting a PGH<sub>2</sub>-EA metabolite in a  
20 sample of the system. In certain embodiments, detection or measurement of a PGH<sub>2</sub>-EA metabolite is performed wherein the metabolite is PGB<sub>2</sub>-EA, PGD<sub>2</sub>-EA, PGE<sub>2</sub>-EA, PGF<sub>2</sub> $\alpha$ -EA, TxB<sub>2</sub>-EA, 6-keto-PGF<sub>1</sub> $\alpha$ -EA, 15-keto-PGE<sub>2</sub>-EA, 13,14-dihydro-15-keto-PGE<sub>2</sub>-EA, PGG<sub>2</sub>-EA, PGH<sub>2</sub>-EA, PGA<sub>2</sub>-EA, PGJ<sub>2</sub>-EA, PGJ<sub>2</sub>-EA derivatives, bicyclo-PGE<sub>2</sub>-EA, TxA<sub>2</sub>-EA or PGI<sub>2</sub>-EA. The presence of the PGH<sub>2</sub>-EA metabolite in  
25 the sample is indicative of the COX-2 activity in the system or subject.

In certain embodiments of the present invention, the inventors provide standards for determining a relative or absolute measurement of COX-2 activity by comparison to a standard or a standard curve generated using the standard. For example, a standard can be used to generate a standard curve for normalization of particular test results. The present invention provides standards including compounds related to reaction products derived from COX-2 action on AEA incorporating isotopic (*e.g.*, radioactive), fluorescent, enzymatic labels and the like.

In certain embodiments, a determination of the amount of COX-2 activity, by the measurement of PGH<sub>2</sub>-EA metabolites, is used to detect inflammation or cancer. In certain embodiments, a relative or absolute determination of inflammation or cancer in the subject is made.

### 3. BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 is a diagram of the chemical structure for arachidonic acid (AA).

FIG. 2 is a diagram of the COX biosynthetic pathway of arachidonic acid to prostaglandins and thromboxane.

FIG. 3 is a diagram of the COX biosynthetic pathway of arachidonic acid to PGH<sub>2</sub>, showing the chemical structure of the substrate and the metabolites.

FIG. 4 is a diagram of the chemical structure for arachidonylethanolamide.

FIG. 5 is a general diagram of enzymatic oxidation of prostaglandin (or thromboxane) ethanolamide (1) and enzymatic hydrolysis of prostaglandin (or thromboxane) ethanolamide (2).

FIG. 6 is a diagram of certain metabolic pathways of PGE<sub>2</sub>-EA illustrating a selection of the type-specific enzymatic and nonenzymatic biotransformations that prostaglandin ethanolamides (PG-EAs) and thromboxane ethanolamides (Tx-EAs) undergo. This diagram of PGE<sub>2</sub>-EA biotransformations can be similarly constructed

for other PG-EAs and Tx-EAs (see, FIGs. 7 and 8). Also, prostaglandin and thromboxane biotransformation pathways can be used as a pattern by one of ordinary skill in the art for the same purpose, in light of the present invention.

FIG. 7 is a diagram of the prostaglandins enzymatically generated following COX-2 action on AEA and in the presence of enzymes downstream of COX-2 (for instance, certain *in vivo* environments).

FIG. 8 is a diagram of certain prostaglandins generated following COX-2 action on AEA *in vitro*.

FIG. 9 is a diagram of tetradeuterated prostaglandin or thromboxane ethanolamide standards.

FIG. 10 is a graph showing the recovery of PGE<sub>2</sub>-EA from human urine.

FIG. 11 is a graph showing the stability of PGE<sub>2</sub>-EA incubated for 5 hours at 37°C in bovine, canine, and human cerebrospinal fluid.

FIG. 12 (A) is a graph showing the stability of PGE<sub>2</sub>-EA in plasma. The PGE<sub>2</sub>-EA (4 mg/mL) remaining after incubation in rat or human plasma for 5 h at 37° C (mean ± S.E., n<sup>3</sup> = 3). (B) PGE<sub>2</sub>-EA (4 mg/mL) is incubated in rat plasma for the indicated times and subjected to LC/MS analysis with selected ion monitoring for the mass corresponding to the dehydration product of PGE<sub>2</sub>-EA (m/z 400). Chromatograms are normalized to the 2 minute sample. (C) PGE<sub>2</sub>-EA (4 mg/mL) are treated as indicated and samples are analyzed for the dehydration product by LC/MS (m/z 400). Chromatograms are normalized to the base treated sample (lowest panel). (D) Is a graph of the absorption spectra of PGE<sub>2</sub>-EA (20 mg/mL) incubated in rat plasma for the indicated times.

FIG. 13 is a graph showing the *in vivo* pharmacokinetics of PGE<sub>2</sub>-EA in a rat given 2 mg/kg PGE<sub>2</sub>-EA by IV. The calculated plasma half-life of PGE<sub>2</sub>-EA is 380 ± 50 seconds (n = 3). A large apparent volume of distribution was observed (300-400 mL)

and both PGE<sub>2</sub>-EA and 13,14-dihydro-15-keto-PGE<sub>2</sub>-EA were routinely detected at 2 hours post-dosing.

#### 4. DETAILED DESCRIPTION OF THE INVENTION

5       The present invention provides novel compositions, methods and articles of manufacture for detecting and measuring cyclooxygenase-2 (COX-2) activity in a subject or a sample thereof. Certain aspects of the present invention focus on detecting and measuring metabolites of AEA, such as, but not limited to, PGE<sub>2</sub>-EA, PGD<sub>2</sub>-EA and TxA<sub>2</sub>-EA. The present invention provides novel compositions, methods and kits for detecting and measuring COX-2 activity, methods for identifying tumors in a subject, evaluating relative tumor severity, and following tumor response to therapy, and methods for detecting inflammation in a subject and evaluating relative inflammation severity. No aspect, embodiment or element, including the claims, of the present invention is bound by theory or mechanism.

##### 4.1 DEFINITIONS

10       In case of conflict, the present document, including definitions, will control. Unless otherwise indicated, materials, methods and examples described herein are illustrative only and not intended to be limiting. All references, citations, articles, publications, patents, and the like provided in this patent application are incorporated herein by reference, in their entirety.

15       The term *in vivo* includes the meaning of processes occurring in an animal, in tissue or cell culture, or in samples taken from an animal or culture.

20       The term *in vitro* includes the meaning of processes occurring in systems wholly or partially purified from the natural environment, such as with purified enzymes or defined enzyme systems.



Purified means partially or wholly isolated away from the natural milieu of factors normally associated with a particular macromolecular species. In certain embodiments, the purified factor comprises 50 percent or more (on a molar basis) of all macromolecular species present in the isolated form. In certain embodiments, a purified composition will comprise more than about 80 percent of all macromolecular species present. In certain preferred embodiments, a purified composition comprises more than about 90 percent of all macromolecular species present. Most preferably, the object species is purified to essential homogeneity (contaminant species cannot be detected in the composition by conventional detection methods) wherein the composition consists essentially of a single macromolecular species. Solvent species, small molecules (<500 Daltons), and elemental ion species are not considered macromolecular species. In non-liquid compositions, "purified" is based upon dry weight and the same percent purities stated above are embodied.

A COX-2 selective substrate is a substrate that is transformed to an enzymatic reaction product by the COX-2 enzyme; but is not transformed, or is not significantly transformed, to a reaction product by the COX-1 enzyme. It is most preferred that a COX-2 selective substrate of the present invention is not enzymatically transformed to a reaction product by COX-1. In certain embodiments, COX-1 may have some activity on the COX-2 selective substrate, but it is not significant relative to the COX-2 activity. Relatively insignificant activity can be determined, for example, by measuring the ratio of substrate oxygenation using purified COX-1 and COX-2.

In certain embodiments, the ratio of COX-1 activity *versus* COX-2 activity for a COX-2 selective substrate, expressed as a percentage, is about 50% or less; in certain embodiments, 40% or less; in certain embodiments, 30% or less; in certain embodiments, 25% or less; in certain embodiments, 20% or less; in certain

embodiments, 10% or less; in certain embodiments, 5% or less; in certain  
embodiments, 3% or less; in certain embodiments, 2% or less; and in certain  
preferred embodiments 1% or less. The lower the percentage (above), the more  
preferred the embodiment. A highly preferred COX-2 selective substrate is  
5 metabolized by COX-2, but is not metabolized by COX-1.

The terms "COX-2 specific substrate" and "COX-2 selective substrate" are  
used interchangeably herein.

In general, enzyme activity refers to the rate at which substrate is consumed  
or product is formed in an enzymatic reaction under a given set of reaction  
conditions. The Standard International (SI) unit for enzyme activity is an enzyme  
unit (U) and is defined as the amount of enzyme needed to produce 1  $\mu$ mole  
product/minute. A unit may be defined differently herein (e.g., the amount of  
enzyme needed to produce 1 nmole product per minute or the amount of enzyme  
needed to consume 1  $\mu$ mole substrate per minute). Additional determinations of  
enzyme activity can be compared when utilizing similar or preferably identical  
reaction conditions. It is understood that reaction conditions can be changed and a  
new enzyme activity scale determined (e.g., by generating a standard curve of  
enzyme activity and use thereof, a process which is known to one of ordinary skill in  
the art). The specific activity of a particular enzyme preparation refers to the total  
20 enzyme units divided by the total amount of protein present in the preparation. A  
preferred unit of specific activity is U per mg of protein (U/mg).

As used herein, references to COX include both COX-1 and COX-2.

Arachidonyl ethanolamide (AEA) is defined herein to be a COX-2 selective  
substrate.

25 The terms "arachidonyl ethanolamide" and "anandamide" are used  
interchangeably herein.

As used herein, "prostaglandin ethanolamides" (PG-EAs) are included in the meaning of COX-2 selective metabolites.

Certain abbreviations include: cyclooxygenase (COX), cyclooxygenase-1 (COX-1), cyclooxygenase-2 (COX-2), prostaglandin (PG), prostaglandins (PGs), ethanolamide (EA), prostaglandin ethanolamides (PG-EAs), arachidonyl ethanolamide (AEA), prostaglandin-ethanolamide (PG-EA), thromboxane (Tx).

As defined herein and using the abbreviations above, PG-EAs can include, but are not limited to: PGB<sub>2</sub>-EA, PGD<sub>2</sub>-EA, PGE<sub>2</sub>-EA, PGF<sub>2</sub>α-EA, TxB<sub>2</sub>-EA, 6-keto-PGF<sub>1</sub>α-EA, 15-keto-PGE<sub>2</sub>-EA, 13,14-dihydro-15-keto-PGE<sub>2</sub>-EA, PGG<sub>2</sub>-EA, PGH<sub>2</sub>-EA, PGA<sub>2</sub>-EA, PGJ<sub>2</sub>-EA, PGJ<sub>2</sub>-EA derivatives, bicyclo-PGE<sub>2</sub>-EA, TxA<sub>2</sub>-EA and PGI<sub>2</sub>-EA (also referred to herein as prostacyclin-EA).

Tumor type typically references the tissue of tumor origin, but can also refer to the current tissue in which a tumor is located (e.g., colon cancer, liver cancer, or pancreatic cancer). The stage and grade of a tumor is related to severity and medical definitions of stages and grades within a continuum are known in the art for each tumor or cancer type. Each specialty within oncology (e.g., hematology, colorectal, liver, pancreatic, lung, brain, dermatology, etc.) may have a particular standard for the stage and grade scale of the tumors used within that clinical specialty, known to one of skill in that art, which varies from the general definitions of tumor stage and grade provided below.

Tumor grade is determined by the appearance of the tumor under the microscope and how quickly the tumor is likely to grow and spread. In general, grading systems are different for each type of cancer, but are known to one of ordinary skill in the art. For example, grade I tumors are the least malignant appearing, grade II tumors are moderately differentiated with a moderately malignant appearance, grade III tumors are less differentiated and show enhanced signs of tissue invasion, and grade IV tumors display the least differentiation and

are the most malignant appearing. The grade of a tumor is determined by one of ordinary skill in the art.

5 The stage of a tumor refers to the extent of a cancer, how advanced the tumor is in the patient (e.g., whether the disease has spread from the original site to other parts of the body). The stage of a tumor is generally determined by radiographic studies such as a computed tomography (CT) scan, magnetic resonance (MRI) imaging and/or ultrasound. Tumor staging is determined by one of ordinary skill in the art and can vary by tumor type or as a field advances, standard staging practices may change. Certain definitions of stages for various cancers are provided in the Dictionary of Cancer Terms on CANCERNET which is a service of the National Cancer Institute available on the World Wide Web at "http://cancernet.nci.nih.gov/dictionary/dictionaryfull.html", incorporated herein by reference in its entirety. A physical location for inquiry or obtaining a copy of the Dictionary of Cancer Terms is: NCI Public Inquiries Office; Building 31, Room 10A03; 31 Center Drive, MSC 2580; Bethesda, MD 20892-2580.

15 Staging refers to performing exams and tests to learn the extent of the cancer within the body, especially whether the disease has spread from the original site to other parts of the body.

## 20 4.2 CYCLOOXYGENASES AND PROSTAGLANDINS

Cyclooxygenase (COX; prostaglandin endoperoxide synthase, EC 1.14.99.1) catalyzes the *bis*-dioxygenation of arachidonic acid (FIG. 1) generating prostaglandin (PG) H<sub>2</sub> (FIGS. 2-3). This is the committed step in prostaglandin and thromboxane biosynthesis. Two isoforms of COX have been cloned from animal cells including constitutively expressed COX-1 (DeWitt, D. L., and Smith, W. L. 1988, *Proc. Natl. Acad. Sci. USA*, 85:1412-1416; Merlie, *et al.* 1988, *J. Biol. Chem.*, 263:3550-3553; Yokoyama, *et al.* 1988, *FEBS Lett.*, 231:347-351; DeWitt, *et al.* 1990,

5 *J. Biol. Chem.*, 265:5192-5198; and Yokoyama, C. and Tanabe, T. 1989, *Biochem. Biophys. Res. Commun.*, 165:888-894) and inducibly expressed COX-2 (Xie, *et al.* 1991, *Proc. Natl. Acad. Sci. USA*, 88:2692-2696; Kujubu, *et al.* 1991, *J. Biol. Chem.*, 266:12866-12872; O'Banion, *et al.* 1991, *J. Biol. Chem.*, 266:23261-23267; Hla, T. and Nielson, K. 1992, *Proc. Natl. Acad. Sci. USA*, 89:7384-7388; Jones, *et al.* 1993, *J. Biol. Chem.*, 268:9049-9054; and Feng, *et al.* 1993, *Arch. Biochem. Biophys.*, 307:361-368).

10 Prostaglandins produced as a result of the activity of COX are known to have numerous physiological functions. These functions include the antithrombogenic action of prostacyclin released by the vascular endothelium and the cytoprotective effect of prostaglandins produced by the gastric mucosa (Whittle, *et al.* 1980, *Nature*, 284:271-273). COX-2 is typically expressed following the activation of normal cells and certain atypically proliferating cells, by various pro-inflammatory agents including certain cytokines (Hla, T. and Nielson, K. 1992, *Proc. Natl. Acad. Sci. USA*, 89:7384-7388; Feng, *et al.* 1993, *Arch. Biochem. Biophys.*, 307:361-368), endotoxin (Lee, *et al.* 1992, *J. Biol. Chem.*, 267:25934-25938) and certain mitogens (Kujubu, *et al.* 1991, *J. Biol. Chem.*, 266:12866-12872; O'Banion, *et al.* 1991, *J. Biol. Chem.*, 266:23261-23267; and Hla, T. and Nielson, K. 1992, *Proc. Natl. Acad. Sci. USA*, 89:7384-7388).

20 Prostaglandins represent a class of substances produced in a wide variety of cells. In general, PGs act on the cells that produce them, on neighboring cells, or over short distances and can be classified as autocrine hormones. PGs and their relatives are usually thought of as potent local hormones (autocrine and paracrine) acting over a short lifetime. PGs, and related compounds, prostacyclin (PGI<sub>2</sub>),  
25 thromboxanes (TX), leukotrienes (LT), and lipoxins (LP), derive from fatty acids stored in cellular membranes as phospholipids or triglycerides, especially arachidonic acid, with an open chain, 20-carbon structure (FIGS. 1-3).

Prostaglandins generally resemble hairpins structurally with a five-membered ring and two chains extending from the ring. In general, substituents on the five-membered ring determine the subclass and activity of the prostaglandins. A series of synthetic reactions catalyzed by enzymes in the membranes, and certain non-enzymatic transformations, culminate in the release of prostaglandin product.

#### 4.3 DETECTING COX-2 ACTIVITY BY DETECTING PROSTAGLANDIN ETHANOLAMIDES

AEA is an unique substrate specific for COX-2 (FIG. 4). COX-2 catalyzes the conversion of AEA to prostaglandin H<sub>2</sub> ethanolamide (PGH<sub>2</sub>-EA) (see FIG. 5-8; Yu, M., et al 1997, *J. Biol. Chem.* 272:21181-21186, incorporated herein by reference). The PGH<sub>2</sub>-EA is subsequently enzymatically and nonenzymatically metabolized to a variety of compounds, such prostaglandin E<sub>2</sub> ethanolamide (PGE<sub>2</sub>-EA) and prostaglandin D<sub>2</sub> ethanolamide (PGD<sub>2</sub>-EA). These downstream metabolites will be referred to, herein, as PGH<sub>2</sub>-EA metabolites. PGH<sub>2</sub>-EA metabolites include, but are not limited to, PGB<sub>2</sub>-EA, PGD<sub>2</sub>-EA, PGE<sub>2</sub>-EA, PGF<sub>2</sub> $\alpha$ -EA, TxB<sub>2</sub>-EA, 6-keto-PGF<sub>1</sub> $\alpha$ -EA, 15-keto-PGE<sub>2</sub>-EA, 13,14-dihydro-15-keto-PGE<sub>2</sub>-EA, PGG<sub>2</sub>-EA, PGH<sub>2</sub>-EA, PGA<sub>2</sub>-EA, PGJ<sub>2</sub>-EA, PGJ<sub>2</sub>-EA derivatives, bicyclo-PGE<sub>2</sub>-EA, HETEs, TxA<sub>2</sub>-EA and PGI<sub>2</sub>-EA., which are metabolized directly from AEA by COX-2, will also be referred to as PGH<sub>2</sub>-EA metabolites in this application. In general, all prostaglandin and thromboxane ethanolamides are susceptible to enzymatic oxidation of the ethanolamide moiety and can undergo oxidation (FIG. 5). A representative example for PGE<sub>2</sub>-EA is shown in FIG. 6. In addition, individual metabolites can enter the 15-hydroxyprostaglandin dehydrogenase/ $\Delta$ <sup>13</sup>-15-keto-prostaglandin reductase pathway to generate 13,14-dihydro-15-keto metabolites, for example 15-keto PGA<sub>2</sub>-EA and 13,14-dihydro-PGA<sub>2</sub>-EA. In general, PGE<sub>2</sub>-EA metabolites can undergo dehydration to enter the PGA<sub>2</sub>-EA/PGB<sub>2</sub>-EA pathway. For example, 13,14-dihydro-

15-ketoPGE<sub>2</sub>-EA can dehydrate to yield 13,14-dihydro-15-keto-PGA<sub>2</sub>-EA and 13,14,-  
dihydro-15-keto-PGB<sub>2</sub>-EA. Finally, oxidative transformation by P450-mediated ω-  
oxidation (e.g., C-19, C-20) can occur.

It has been shown that AEA is found endogenously (Devane *et al.* 1992,  
5 *Science* 258:1946-1949, hereby incorporated by reference). The amount of PGH<sub>2</sub>-EA  
metabolites in a biological sample correlates with the COX-2 specific activity in the  
subject from which the sample was collected. The amount of COX-2 activity is a  
marker of or a measurement of inflammatory or cancerous disease processes. Certain  
embodiments of the present invention include methods and materials for making  
10 sensitive measurements of picogram quantities of PGH<sub>2</sub>-EA metabolites. Further  
embodiments of the present invention include using these measurements of COX-2  
activity to clinically grade or stage a disease process and to assess therapeutic  
outcomes.

#### 15 4.4 MEASURING PROSTAGLANDIN ETHANOLAMIDES

For the purposes of this application, detecting includes determining if a  
substance or compound is present in a sample. In the present invention, the substance  
or compound being detected is a PGH<sub>2</sub>-EA metabolite. Detecting can include  
measuring. In general, measuring means determining the relative or absolute amount  
20 of a substance or compound detected. Measurement is generally, but not always,  
performed relative to a standard. For example, the amount of the standard may be  
correlated with an amount of COX-2 activity or expression. Therefore, comparing the  
amount of PGH<sub>2</sub>-EA metabolites measured in a sample from a subject indicates an  
amount of COX-2 activity or expression in the subject.

25 PGH<sub>2</sub>-EA metabolites may be detected and measured in a variety of ways  
(see Examples 1-6). In certain embodiments of the present invention, for example, a  
sample is collected from a subject, and a selective COX-2 substrate, such as AEA, is

added. Then, a metabolite of the COX-2 specific PGH<sub>2</sub>-EA product is measured. In preferred embodiments, downstream metabolites are measured because the half-life of PGH<sub>2</sub>-EA in aqueous solution is approximately 20 seconds. Certain downstream metabolites are more stable and are preferred for measurement. A highly preferred downstream metabolite for measurement is PGE<sub>2</sub>-EA. In another embodiment of the present invention, for example, both a COX-1 substrate (such as arachidonic acid) and a COX-2 selective substrate (such as AEA) are added to the sample. Then the amount of downstream metabolites derived from the enzymatic actions of COX-1 and COX-2 on each substrate are measured and compared. In both examples, the samples can be incubated with the substrates over time and a series of measurements of metabolites taken and compared in relation to the amount of time that passed. In certain embodiments of the present invention, no substrate is added to the sample. Instead, the PGH<sub>2</sub>-EA metabolites derived from the endogenous AEA is detected and measured. A highly preferred downstream metabolite for measurement is 13,14-dihydro-15-keto-PGE<sub>2</sub>-EA, a novel lipid formed by the sequential oxidation and reduction of PGE<sub>2</sub>-EA by 15-hydroxyprostaglandin dehydrogenase and  $\Delta$ 13,15-keto-prostaglandin reductase. This metabolite is formed *in vivo* following intravenous dosing of PGE<sub>2</sub>-EA to rats and is detectable in plasma by the methods described herein. In some cases, AEA is administered to the subject prior to sample collection, followed by sample collection, and detection and measurement of PGH<sub>2</sub>-EA metabolites present in the sample. In addition, in preferred embodiments, the amount of PGH<sub>2</sub>-EA metabolites measured in the sample is related to an amount of COX-2 activity in the sample or the subject.

Samples from the subject can be processed is several ways (see Examples 5-6). For example, the sample may be extracted at least one time with a solvent, to remove the PGH<sub>2</sub>-EA metabolites from the sample for analysis. Extraction can be



followed by evaporation. The resulting residue may be redissolved in another solvent and analyzed. This process might involve several rounds of the extraction, evaporation and redissolving steps. Alternatively, the solution resulting from one or more extractions of the sample may be filtered and analyzed. In certain preferred  
5     embodiments, the sample is extracted, filtered and analyzed for PGH<sub>2</sub>-EA metabolite content.

One aspect of the present invention is a method of detecting COX-2 activity in a biological sample, comprising: incubation of the biological sample with AEA, extracting the sample with a solvent, evaporating the solvent to leave a residue and analyzing the residue for PGH<sub>2</sub>-EA metabolites wherein the presence of PGH<sub>2</sub>-EA metabolites is indicative of COX-2 activity (for instance, see Examples 1, 2, and 3). In certain embodiments, the amount of PGH<sub>2</sub>-EA metabolites is measured and related to the quantity of COX-2, COX-2 expression, or COX-2 activity. In general, the analysis of PGH<sub>2</sub>-EA metabolites includes, but is not limited to, detection by liquid chromatography- mass spectrometry (Examples 1-2, 5-6).

Another embodiment of the present invention is a method of detecting COX-2 activity in a biological sample, comprising: extracting PGH<sub>2</sub>-EA metabolites present in the sample with a solvent, evaporating the solvent to leave a residue and quantifying PGH<sub>2</sub>-EA metabolites in the residue, wherein the presence of PGH<sub>2</sub>-EA  
20     metabolites is indicative of COX-2 activity. Preferably, the quantity of PGH<sub>2</sub>-EA metabolites detected are related to a quantity of COX-2 activity.

Yet another embodiment of the present invention is a method of detecting a COX-2 activity in a biological sample, comprising: extracting the sample at least one time with a first solvent, evaporating the solvent to leave a residue, dissolving the  
25     residue in a second solvent, separating at least one PGH<sub>2</sub>-EA metabolite from the dissolved residue with a separation device, lyophilizing the separated PGH<sub>2</sub>-EA

metabolite, dissolving the lyophilized PGH<sub>2</sub>-EA metabolite in a third solvent and detecting the dissolved PGH<sub>2</sub>-EA metabolite with a detection device, wherein the presence of the PGH<sub>2</sub>-EA metabolite is indicative of the COX-2 activity (for instance, see Examples 1, 2, and 3). In certain embodiments, the amount of PGH<sub>2</sub>-EA metabolite is measured and related to a quantity of COX-2 protein, COX-2 expression, or COX-2 activity. In general, the separation device for separating the PGH<sub>2</sub>-EA metabolite from a residue in the second solvent includes, but is not limited to, a liquid chromatography device.

Yet another aspect of the present invention is a method of detecting COX-2 activity in a biological sample, comprising: extracting at least one PGH<sub>2</sub>-EA metabolite from the sample with a solvent, evaporating the solvent to leave a residue, dissolving the residue in a second solvent, filtering the dissolved residue and detecting a PGH<sub>2</sub>-EA metabolite in the filtered solution with a detection device, wherein the presence of PGH<sub>2</sub>-EA metabolite in the sample is indicative of COX-2 activity. Preferably, the detected PGH<sub>2</sub>-EA metabolite is measured and the amount of PGH<sub>2</sub>-EA metabolite measured is related to a quantity of COX-2 activity.

#### 4.4.1 USE OF A STANDARD

In certain embodiments, isotopically labeled PGH<sub>2</sub>-EA metabolites, such as PGE<sub>2</sub>-EA and TxA<sub>2</sub>-EA, are used as an internal standard in quantifying PG-EAs. In certain preferred embodiments, mass spectrometric quantification of PGH<sub>2</sub>-EA metabolites is performed. One example of the synthesis of isotopically labeled ethanolamides of prostaglandins and thromboxanes is shown in FIG. 9. In general, isotopically labeled PG- or Tx-EA are prepared by coupling the target PG or Tx free acid with anhydrous, isotopically labeled ethanolamide and purified by chromatography. In another example, labeled standard ethanolamides of

prostaglandins and thromboxanes can be labeled by substituting  $^2\text{H}$  or  $^3\text{H}$  for a  $^1\text{H}$  attached to at least one of the carbons 2-20 of the arachidonyl carbon chain. Alternatively, a  $\text{PGH}_2\text{-EA}$  metabolite can be labeled by substituting  $^{13}\text{C}$  or  $^{14}\text{C}$  for at least one  $^{12}\text{C}$  in the arachidonyl carbon chain. Labeled  $\text{PGH}_2\text{-EA}$  metabolites can also be made by reacting labeled AEA with COX-2, followed by purification of the reaction products. In general, labeled  $\text{PGH}_2\text{-EA}$  metabolites may also be synthesized by substituting at least one atom in the molecule with an isotope. In certain embodiments, the isotope is non-positron emitting. In certain embodiments, the isotope is positron emitting. Alternatively,  $\text{PGH}_2\text{-EA}$  metabolites can be derivatized to produce chemiluminescent or fluorescent standards, which can be used, for example, in  $\text{PGH}_2\text{-EA}$  quantitation using high performance liquid chromatographic separation coupled with fluorescence or chemiluminescence detectors.

Labeled PG-EA or Tx-EA standard curves, which correlate with the presence or absence of cancer in a patient, can be generated. For example, this can be accomplished by collecting samples from groups of patients with different types, stages, or grades of cancer, measuring the amount of a  $\text{PGH}_2\text{-EA}$  metabolites in the samples, and graphing the amounts of the  $\text{PGH}_2\text{-EA}$  metabolite *versus* the types of cancer. In certain embodiments, types can be varieties. In preferred embodiments, the types are a range of particular cancers measured on a clinical scale of severity or aggressiveness. Such a standard curve may indicate that the larger the amount of cancer or the more progressed the cancer in the patient, the more  $\text{PGH}_2\text{-EA}$  metabolite found in the sample from the patient. In addition, since small amounts of  $\text{PGH}_2\text{-EA}$  metabolites can be found in samples collected from people without cancer, a threshold level of  $\text{PGH}_2\text{-EA}$  metabolites will be observed at which amounts of  $\text{PGH}_2\text{-EA}$  metabolites below the threshold will correlate with the lack of cancer in the patient. On the other hand, for example, amounts of  $\text{PGH}_2\text{-EA}$  metabolites

measured in a sample that are above the threshold level correlate with the presence of cancer in the patient.

In a similar example, a standard curve can be developed which correlates the amount of PGH<sub>2</sub>-EA metabolites observed in a sample from a patient with an amount of inflammation. Such a standard curve can be generated by collecting samples from a patient population suffering from inflammatory processes, and correlating the amounts of PGH<sub>2</sub>-EA metabolite in the samples with the severity of disease in the patient population.

In another example, no standard or standard curve is used. Instead, a series of samples may be collected from a single patient over a period of time, and the amount of PGH<sub>2</sub>-EA metabolite in each sample measured. Then the amounts of PGH<sub>2</sub>-EA metabolite measured would be compared and correlated with the amount of time which had passed between sample collections. If the amount of PGH<sub>2</sub>-EA metabolite in the samples increased over time, for example, this would indicate that the patient's disease state is worsening, or that therapeutic intervention is not effective. On the other hand, if the amount of PGH<sub>2</sub>-EA metabolite measured in the samples decreased over time, for example, this would indicate that the patient's disease state is improving or that therapeutic intervention is successful.

#### **4.4.2 DETECTION AND MEASURING DEVICE**

In general, a detection device for detecting PGH<sub>2</sub>-EA metabolites includes, but is not limited to, a mass spectrometer, a chromatography-coupled mass spectrometer, an immunoassay or an enzyme-linked immunoassay, or other means for detecting PGH<sub>2</sub>-EA metabolites known in the art (see Examples 1-6). For example, in certain embodiments of the present invention, liquid chromatography/mass spectrometry (LC/MS) is conducted, preferably with a Waters 2690 Separations Module with a Zorbax RX-C18 narrow bore column (15 cm x 2.1

mm, 5  $\mu$ m) interfaced to a Finnigan TSQ-7000 triple quadrupole mass spectrometer. Sodiased analytes are eluted with increasing concentrations of MeCN in 0.001% aqueous sodium acetate. Evaluation of PGH<sub>2</sub>-EA metabolites in biological samples is conducted with selected ion monitoring and quantification is accomplished using  
5 tetradeutereated standards, such as tetradeutereated PGE<sub>2</sub>-EA. Electrospray ionization (ESI) is carried out using nitrogen as sheath (76 psi) and auxiliary gas (14 psi) to assist with nebulization. A potential of 5.5 kV is applied to the ESI needle and the capillary temperature is maintained at 220 °C. Mass spectral parameters are optimized to obtain maximum sensitivity without sacrificing unit resolution.

#### 4.4.3 SEPARATION DEVICE

A variety of separation devices known in the art for separating prostaglandins from a sample may be used. In general, separation devices include, but are not limited to, extraction columns, affinity columns, filters, thin-layer chromatography plates and gels.

In certain embodiments of the present invention, a PGH<sub>2</sub>-EA metabolite in a sample may be isolated or purified (separated partially or substantially from the natural constituents of a PGH<sub>2</sub>-EA metabolite containing sample) using one or more techniques known in the art for the separation of chemical and especially  
20 prostaglandin compounds, for example, but not limited, to liquid chromatography.

#### 4.4.4 SUBJECTS

In certain embodiments of the present invention, the subject includes a mammal, such as a rodent, preferably a human, or a cultured cell of a mammal,  
25 including a cultured cell of a human. Other subjects include farm animals and show animals (horses, cattle, sheep, pigs and swine, goats, fowl, and the like), pets (dogs, cats, parrots, canaries and the like), animals kept in zoos and endangered species

(elephants, lions, tigers, antelope, zebra, anteaters, water buffalo, pandas, cheetahs, kangaroos, ostriches, eagles, condors, finches, and the like) or a cultured cell of said animal.

#### 5 4.4.5 SAMPLES

10 In certain embodiments, the sample is urine or may be collected from or processed from urine. PGH<sub>2</sub>-EA metabolites can be measured in urine by isotope dilution mass spectrometry. A fixed volume of urine is treated with an appropriate internal standard (*e.g.*, tetradeutereated PGE<sub>2</sub>-EA) and then loaded on reversed-phase extraction cartridges. The sample is then washed (*e.g.*, 1 ml pH 4.0 20 mM sodium acetate) and PGH<sub>2</sub>-EA metabolites, including the added internal standard, are eluted with organic solvent (*e.g.*, two 1 ml aliquots of MeCN). The solvent is evaporated and the residue is analyzed by LC/MS. A typical procedure wherein synthetically generated PGE<sub>2</sub>-EA and *d*5-PGD<sub>2</sub>-EA is added to fresh human urine is depicted in FIG. 10.

15 PGH<sub>2</sub>-EA metabolites are stable in cell culture, bovine, canine and human CSF (FIG. 11) and human and rat plasma (FIG. 12) incubated 5 hours at 37°C. FIG. 13 shows the *in vivo* pharmacokinetics of PGE<sub>2</sub>-EA in a rat given 2 mg/kg PGE<sub>2</sub>-EA intravenously. The calculated half-life of PGE<sub>2</sub>-EA is 380 ± 50 seconds (n=3).  
20 Notably, this half-life greatly exceeds that observed for the free acid PGE<sub>2</sub> *in vivo*, highlighting one significant advantage of the methods described herein over previously described methods for COX activity assessment *in vivo*. A large apparent volume of distribution was observed (300-400 ml) and both PGE<sub>2</sub>-EA and 13,14-dihydro-15-keto- PGE<sub>2</sub>-EA were routinely detected at 2 hours post-dosing. In certain  
25 embodiments, the sample is blood, plasma, cerebrospinal fluid, saliva, sputum, bile, joint fluid, biopsy, immune cells, cancer cells, tumor cells, malignant cells,

inflammatory cells, non-tumor cells, non-immune cells, cells not activated by inflammatory stimuli or may be collected from or processed from one or more of these fluids and tissues. In certain embodiments of the present invention, the sample may be conditioned media from a cell culture. In certain embodiments of the present invention, a plurality of samples may be collected from the subject, with a period of time being allowed to pass between consecutive collections of the samples. The amounts of PGH<sub>2</sub>-EA metabolites present in these samples are measured, compared and related to the periods of time that had been allowed to pass between collections. In certain embodiments, the subject is a patient and the samples are taken in order to evaluate the effectiveness of anti-cancer therapy and to evaluate tumor state, severity or load in the patient.

In general, samples can be collected from or prepared from cultured cells (Example 6). A variety of cells lines known to one skilled in the art are acceptable. In addition, primary cell cultures can be used. Methods for collecting and culturing primary cell cultures are well known in the art.

#### 4.4.6 ANTIBODY SYNTHESIS

Monoclonal and polyclonal antibodies to PGH<sub>2</sub>-EA metabolites or their metabolites can be made using standard antibody generation techniques in light of the present invention (Cohen, et al., U.S. Patent Serial No. 5,589,575, herein incorporated by reference; McCafferty, et al. 1996, *Antibody Engineering, a Practical Approach*, IRL Press; Mernaugh & Mernaugh 1994, *Methods for the Production of Monoclonal Antibodies*, in **Molecular Methods in Plant Pathology**). For example, monoclonal antibodies against PGE<sub>2</sub> are commercially available from Cayman Chemical (118 E. Ellsworth Rd., Ann Arbor, MI 48108, 800-364-9897) and chemiluminescent ELISA kits for several PGs, HETE and TxB<sub>2</sub> are available from Assay Designs, Inc. (800 Technology Dr., Ann Arbor, MI 48108; 734-

668-6113). To create antibodies that recognize PGE<sub>2</sub>-EA, for example, the cyclopentyl substituents and amide moieties of the PGE<sub>2</sub>-EA are protected using standard techniques. Then the protected PGE<sub>2</sub>-EA is chemically linked to an appropriate conjugate (*e.g.*, keyhole limpet hemocyanin (KLH)) preferably *via* 5 covalent attachment at carbon 15. The haptenized product is deprotected and the resultant immunogen is used to inoculate mice, rabbits, or goats using standard techniques. According to standard protocols, serum would be collected from the immunized animals and antibodies are purified from the serum. Alternative antibody development strategies using the same immunogen could also be employed (single-chain antibodies, hybridomas, etc.). 10

#### 4.5 COX-2 AND INFLAMMATORY DISEASES/DISORDERS

A wide variety of human diseases are associated with inflammation. These range from acute appendicitis to asthma, myocardial infarction, specific immunological disease processes, infection with viruses or bacteria, malignancy and metastasis, endotoxemia and reperfusion injury. Since COX-2 activity is important in the progression of these diseases, the present invention is a useful method of diagnosing or monitoring disease state. The present invention is also useful in detecting and treating non-malignant or immunological-related cell-proliferative 20 diseases such as psoriasis, pemphigus vulgaris, Behcet's syndrome, acute respiratory distress syndrome (ARDS), ischemic heart disease, post-dialysis syndrome, leukemia, acquired immune deficiency syndrome, septic shock and other types of acute inflammation, and lipid histiocytosis. Essentially, the present invention can be used to facilitate the detection, measurement and treatment of any 25 disorder which is etiologically linked to the inflammatory process (for instance, see Example 4).



For example, in certain aspects of the present invention, a sample is collected from a patient suspected of having an inflammatory disease or diagnosed with an inflammatory disease and PGE<sub>2</sub>-EA in the sample is detected and measured. The amount of PGE<sub>2</sub>-EA present in the sample correlates with COX-2 activity in the patient and is a marker of the progress or severity of that disease or disease process. In another example, a series of samples are collected from the patient over a period of time. During that period of time, the patient undergoes treatment for the inflammatory disease. Changes in the amount of PGE<sub>2</sub>-EA measured over time are indicative of changes in COX-2 activity and changes in the patients disease state. This information would be used by the physician to evaluate the patient's condition as well as the effectiveness of therapeutic intervention, wherein a decrease in the PGH<sub>2</sub>-EA metabolites or COX-2 activity is indicative of an improvement in the patient's condition or effective therapy.

#### 4.6 COX-2 AND CANCER

Studies in human colon cancer have shown that COX-2 expression is increased in colon cancer cells compared to the adjacent colonic mucosa; similar observations have been made in experimental models of colon cancer (Eberhart, CE, *et al.* 1994, *Gastroenterology* 107:1183; Sheng, H, *et al.* 1997, *J. Clin. Invest* 99:2254; DuBois, RN, *et al.* 1996, *Gastroenterology* 110:1259). COX-2 expression is a marker for the metastatic potential of colon cancer cells and is related to patient survival (Tsuji, M, *et al.* 1997, *Proc. Natl. Acad. Sci. USA* 94:3336; Sheehan, KM, *et al.* 1999, *JAMA* 282:1254). In one study, for example, COX-2 expression was determined in 76 patients with a variety of stages of colorectal cancer (Sheehan, KM, *et al.*, 1999, *JAMA* 282:1254). Such studies can be used to generate a standard curve for COX-2 expression in cancer and colon cancer in particular (see *supra*).

Ten-year survival was significantly higher in patients with the lowest levels of COX-2 expression (68 versus 35 percent). These findings suggest that COX-2 activation promotes tumor growth. Consistent with this hypothesis is a study in which human colon cancer cells that expressed high levels of COX-2 were implanted into nude mice. Treatment with a selective COX-2 inhibitor reduced tumor formation by 85 to 90 percent and inhibited colony formation of cultured cells (Sheng, H, *et al.* 1997, *J. Clin. Invest* 99:2254). This benefit was not seen with tumor cells that lacked COX-2.

Certain aspects of the present invention include methods of detecting a tumor in a patient in need thereof, comprising: obtaining a sample from the patient and detecting at least one PGH<sub>2</sub>-EA metabolite in the sample (for instance, see Example 5). The presence of the PGH<sub>2</sub>-EA metabolite in the sample is a marker for the presence of the tumor in the patient. More preferably, an amount of PGH<sub>2</sub>-EA metabolite detected in the sample of the patient will be measured, wherein the amount of PGH<sub>2</sub>-EA metabolite measured is indicative of the amount or severity of tumor present in the patient.

A further aspect of the present invention is a method of measuring and monitoring the size, grade, and/or stage of a tumor, comprising: collecting a first sample of a subject and measuring the amount of PGH<sub>2</sub>-EA metabolites in the first sample. Then a period of time is allowed to pass, during which the subject may, or may not, undergo anti-cancer therapy. After the period of time has passed, a second sample is collected from the subject and the amount of PGH<sub>2</sub>-EA metabolites in the second sample is measured. The amounts of the PGH<sub>2</sub>-EA metabolites in the first and second samples are compared, wherein the difference between the amounts of PGH<sub>2</sub>-EA metabolites in the two samples is indicative of changes in the metabolism of the cancer cell.

In certain embodiments of the present invention, a plurality of samples may be collected from the subject, with a period of time being allowed to pass between consecutive collections of the samples. The amounts of PGH<sub>2</sub>-EA metabolites present in these samples are measured, compared and related to the periods of time that had been allowed to pass between collections. In certain embodiments, the subject is a patient and the samples are collected in order to evaluate the effectiveness of anti-cancer therapy and to evaluate tumor severity or tumor load in the patient.

In general, the effectiveness of the anti-cancer therapy is evaluated based on the changes in the amount of sample PGH<sub>2</sub>-EA metabolite observed over time. For example, increases in the amount of PGH<sub>2</sub>-EA metabolites over time indicate continued tumor growth and failure of therapeutic intervention; whereas decreases in the amount of PGH<sub>2</sub>-EA metabolites over time are indicative of therapeutic success and tumor regression. In certain embodiments of the present invention, the sample is a culture of cancer cells used as an experimental model or a culture of cancer cells taken from a patient. The cultured cancer cells may be treated with an anti-cancer therapy *in vitro* in order to evaluate the effectiveness of that therapy in relation to alternative cancer therapies. In certain embodiments, this procedure is done in order to determine an optimal anti-cancer therapy for that individual patient.

In certain embodiments, the attending health professional may characterize both an inflammatory process and a malignancy in the subject by detecting or measuring an amount of a prostaglandin ethanolamide in a sample of a subject specifically produced by the offending malignancy and inflammatory lesion.

#### 4.7 COX-2 AND RESEARCH

In the prior art, investigations which attempt to identify links between COX-2 expression/activity and disease processes are time- and labor-intensive and often require examination of tissue samples post-mortem. For example, attempts to study the role of COX-2 in Alzheimer's disease (AD) require post-mortem collection of brain tissue from both deceased AD and control subjects and quantitative assessment of COX-2 expression in this tissue using standard biochemical techniques (e.g., Western blotting). Such studies are also hampered by the inability to assess enzyme activity, which may or may not correlate with enzyme expression. The use of PGH<sub>2</sub>-EA metabolite quantification in this setting allows for a relatively non-invasive quantification of COX-2 activity *in vivo*. This technique provides at least two fundamental benefits. First, the noninvasive nature allows for much broader testing increasing the sample size in these studies and permitting rapid and statistically significant association (or lack thereof) between COX-2 activity and the pathology under study. Second, given the possibility of testing patients before disease signs are evident will allow for assessing the role of COX-2 in disease development and progression in contrast to post-mortem studies which evaluate the role of COX-2 long after the disease process began. Quantification of PGH<sub>2</sub>-EA metabolites *in vivo* provides a simple assay for assessing the *in vivo* efficacy of newly developed COX-2 inhibitors.

In addition, there is a great need to discover and develop new COX-2 specific regulators. However, current methods are time and labor intensive. The present invention provides compositions, methods and kits for screening candidate molecules for their ability to regulate COX-2 using cultured cells, tissues, or whole animals (Examples 9-10).

#### 4.8 KITS

Certain embodiments of the present invention provide an article of manufacture for the detection and/or measurement of COX-2 activity by the detection and/or measurement of PGH<sub>2</sub>-EA metabolites by radioassay or immunoassay, comprising an antibody and a set of instructions delineating a process for relating a detection and/or measurement of PGH<sub>2</sub>-EA metabolites in a sample to a detection and/or measurement of COX-2 in a subject or a sample thereof. Preferably, the article of manufacture further comprises an antibody against PGH<sub>2</sub>-EA metabolites. More preferably, the article of manufacture further comprises the standard reagents required to perform an immunoassay, such as buffers, multi-well plates, additional antibodies and the like. Still more preferably, the article of manufacture further comprises one or more solid phase extraction columns for the isolation/purification of PGH<sub>2</sub>-EA metabolites. Preferably, the article of manufacture further comprises a set of standards. More preferably, the article of manufacture further comprises an unlabeled PGH<sub>2</sub>-EA metabolite internal standard for standard curve development.

In further embodiments the present invention provides an article of manufacture for the detection and/or measurement of COX-2 activity by mass spectrometry, which comprises: a set of instructions delineating a process for relating a detection and/or measurement of PGH<sub>2</sub>-EA metabolites in a sample to a detection and/or measurement of COX-2 in a subject or a sample thereof and a C18 solid phase extraction column. Preferably, the article of manufacture further comprises a set of standards. More preferably, the article of manufacture further comprises an unlabeled PGH<sub>2</sub>-EA metabolite positive control, and a tetradeuterated PGH<sub>2</sub>-EA metabolite internal standard.

## 5.0 EXAMPLES

### 5.1 Example 1

In general, COX-2 enzymatic products can be detected or measured by a variety of methods. For example, reversed phase liquid chromatography-mass spectrometry is used to detect or measure COX-2 enzymatic products. Selected-ion mass chromatograms of sodium adducts of oxygenated AEA metabolites can be generated by monitoring  $m/z = 386$  (HETE-EAs), 418 (PG-EAs), and 436 (Tx-EA and 6-keto-PGF $1\alpha$ -EA). For example, products are eluted with a 15-minute gradient of 20% to 100% acetonitrile and H<sub>2</sub>O (0.001% sodium acetate). LC/MS reveals the presence of five primary products in AEA/COX-2 reaction mixtures. Two closely eluting polar products each display  $m/z$  418 consistent with the non-enzymatic PGH<sub>2</sub>-EA isomerization products PGE<sub>2</sub>-EA and PGD<sub>2</sub>-EA. Tetradeutereated standards of PGE<sub>2</sub>-EA and PGD<sub>2</sub>-EA are synthesized and coeluted with these two polar products under multiple chromatographic conditions. An intermediate polarity product with  $m/z$  346 is observed that is consistent with the PGH<sub>2</sub>-EA degradation product HHT-EA. Finally, two closely eluting non-polar AEA metabolites are detected with an  $m/z$  of 386, consistent with the ethanolamides of HETEs (hydroxy-eicosatetraenoic acids that are minor products of arachidonic acid oxygenation by COX-2).

### 5.2 Example 2

Cellular COX-2 enzymatic activity can be determined by exposing cells to exogenous AEA and measuring the COX-2 specific products. For example, unactivated RAW264.7 cells (a murine macrophage cell line) express no detectable COX-2 and low levels of COX-1, but IFN- $\gamma$  and LPS induce COX-2 expression (Wadleigh, D. J. et al, 2000, *J. Biol. Chem.* 275:6259-6266). The major arachidonic

acid metabolite in these cells is PGD<sub>2</sub>, so RAW264.7 cells permit the simultaneous evaluation of AEA oxygenation to PGH<sub>2</sub>-EA as well as endoperoxide metabolism by PGD synthase (Landino, L. M., Crews, B. C., Timmons, M. D. *et al.*, 1996, *Proc. Natl. Acad. Sci. USA* 93:15069-15074). PGH<sub>2</sub>-EA metabolites are not detectable in  
5 the medium from unactivated RAW264.7 cells following addition of AEA (20 μM) (Kozak *et al.* 2000, *supra*). However, treating cells with LPS (1 μg/mL) and IFN-γ (10 units/mL) results in the synthesis and release of copious amounts of PGD<sub>2</sub>-EA, following the addition of AEA (20μM). A LC-mass spectrogram showing the PGD<sub>2</sub>-EA production and extracellular release by activated RAW264.7 macrophages can be generated. Products are eluted with a 15-minute gradient of 20% to 100% acetonitrile in H<sub>2</sub>O (0.001% sodium acetate). Chromatograms are normalized to total ion current of AEA- treated activated macrophages. PGD<sub>2</sub>-EA biosynthesis is inhibited both by indomethacin (3 μM) and the highly selective COX-2 inhibitor, 2-[1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl]-*N*-phenethylacetamide  
10 (indomethacin phenethylamide, 3 μM). No significant endogenous PGD<sub>2</sub>-EA biosynthesis is detected in the absence of exogenous substrate under these conditions.

### 20 5.3 Example 3

A sample may be urine or may be collected from or processed from urine. PGH<sub>2</sub>-EA metabolites can be measured in urine by isotope dilution mass spectrometry. A fixed volume of urine is treated with an appropriate internal standard (*e.g.*, tetradeutereated PGE<sub>2</sub>-EA) and then loaded on reversed-phase  
25 extraction cartridges. The sample is then washed (*e.g.*, 1 ml pH 4.0 20 mM sodium acetate) and PGH<sub>2</sub>-EA metabolites including the added internal standard are eluted with organic solvent (*e.g.*, two 1 ml aliquots of MeCN). The solvent is evaporated

and the residue is analyzed by LC/MS. A typical procedure wherein synthetically generated PGE<sub>2</sub>-EA and *d4*-PGD<sub>2</sub>-EA is added to fresh human urine is depicted in FIG. 10 and demonstrates that this technique provides precise and linear quantitation over the tested range (0-40 ng PGE<sub>2</sub>-EA per ml urine).

5

#### 5.4 Example 4

COX-2 expression and activity are generally linked with the inflammatory process, which accompanies a plethora of pathologies including, but not limited to, arthritis/arthropathy, infectious disease, neurodegenerative disease, neoplasia and autoimmune disease. The quantification of prostaglandin PGH<sub>2</sub>-EA metabolites from biological fluids obtained non-invasively (*e.g.*, blood, urine) will allow for the assessment of COX-2 activity *in vivo*, reflecting both inflammation and disease severity. In addition, serial testing will allow for the tracking of the natural course of the disease as well as the efficacy of anti-inflammatory therapy. A model for this application would be the ubiquitous use of C-reactive protein (CRP) in the diagnosis and assessment of diseases associated with inflammation. The benefits of using PGH<sub>2</sub>-EA metabolites in this context instead of more traditional diagnostic markers, such as CRP, involve the highly specific nature of PGH<sub>2</sub>-EA metabolite production. PGH<sub>2</sub>-EA metabolites are elevated only when COX-2 activity is elevated whereas CRP elevations, for example, are very non-specific.

10  
15  
20

**Scenario:** Elderly woman seeks medical attention for recent onset of joint pain in hands. Urine is collected and PGH<sub>2</sub>-EA metabolite quantification is conducted. PGH<sub>2</sub>-EA metabolites are elevated supporting a diagnosis of rheumatoid arthritis. COX-2 inhibitor therapy is initiated (*e.g.*, celecoxib). After one week, symptoms are only mildly relieved and another urine sample reveals PGH<sub>2</sub>-EA metabolite levels are still elevated, indicating that the inflammatory process is still

25



active. Following dosage increase, symptoms are relieved and urinary PGH<sub>2</sub>-EA metabolites are normalized.

### 5.5 Example 5

5 COX-2 expression and activity are linked to several solid tumors, most notably colorectal adenocarcinoma. The quantification of PGH<sub>2</sub>-EA metabolites from biological fluids, described herein, provides a noninvasive "early-warning" for clinically undetectable neoplasia. In addition, serial testing following diagnosis will allow for the tracking of the natural course of the cancer as well as the efficacy of antineoplastic therapy. A model for this application would be the use of prostate specific antigen (PSA) in the diagnosis and assessment of prostate adenocarcinomas. The benefits of PGH<sub>2</sub>-EA metabolite quantification in this context include (a) relative noninvasiveness, (b) sensitivity (most cancers are advanced once symptomatic) and (c) cost (simple lab diagnostic technique versus colonoscopy for example).

10 **Scenario:** Elderly asymptomatic man receives annual physical examination. Urine and blood are collected and PGH<sub>2</sub>-EA metabolite quantification is conducted. PGH<sub>2</sub>-EA metabolites are elevated in both the urine and plasma, prompting a more detailed search for possible neoplasia. Colonoscopy reveals a single tumor in the descending colon which, following biopsy, proves malignant. Standard chemotherapy is initiated. Following treatment, urinary and plasma PGH<sub>2</sub>-EA metabolite levels have normalized. Annual colonoscopies for 3 years reveal no recurrence. After 3 1/2 years, patient visits physician for an unrelated reason and urinary PGH<sub>2</sub>-EA metabolites are quantified. PGH<sub>2</sub>-EA metabolite levels are markedly elevated, indicating the recurrence of carcinoma. The physician recommends colonoscopy, which reveals the presence of carcinoma. Aggressive chemotherapy is initiated and urinary PGH<sub>2</sub>-EA metabolite levels are monitored.

## 5.6 Example 6

The measurement of PGH<sub>2</sub>-EA metabolites from *in vitro* samples (*e.g.*, cell culture, biopsy samples) allows for the direct quantification of COX-2 activity. Current methods which quantify cyclooxygenase activity do not directly distinguish between COX-1 and COX-2. Methods which quantify COX-2 expression (*e.g.*, Western blotting) do not assess activity which may or may not correlate with expression levels.

**Scenario:** Researchers investigating new NSAIDS expose cultured cells expressing COX-2 to various concentrations of test compounds for predetermined periods of time. At the conclusion of the exposures, conditioned medium is collected from each culture. The samples of conditioned medium are assayed for the presence of PGH<sub>2</sub>-EA metabolite. The researchers find that most of the test compounds have no significant affect on the production of PGH<sub>2</sub>-EA metabolite by the cultured cells. However, one compound ("compound X") dramatically reduces the amount PGH<sub>2</sub>-EA metabolite produced by the cultured cells. Therefore, "compound X" inhibits COX-2. The researchers focus their efforts on "compound X," which may become a new COX-2 specific treatment of inflammatory diseases or cancer. In further experiments, the researchers determine that compound X does not inhibit COX-1. Compound X is, therefore, identified as a COX-2 specific inhibitor.

## 5.7 Example 7

In certain embodiments, the present invention can be employed to identify a previously undescribed small molecule modulator of COX-2 activity. This can be done with the following method. Briefly, RAW264.7 cells at 30-40% confluence are activated with lipopolysaccharide (LPS, 20 ng/mL) and treated with a series of concentrations of a test compound. Cells are incubated for 12 h at 37°C and then

medium is removed and replaced with buffered saline. Cells are then treated with 50  $\mu$ M AEA and incubated an additional 30 min at 37°C. Following incubation, buffered saline is collected and treated with tetradeutereated PGH<sub>2</sub>-EA metabolite standard. Buffered saline is extracted twice with equal volumes of 2:1 CHCl<sub>3</sub>:MeOH. The combined organic extract is evaporated under a stream of argon. The resultant residue is redissolved in 1:1 H<sub>2</sub>O:MeCN and analyzed by liquid chromatography-mass spectrometry (LC-MS) with selected ion monitoring of PGH<sub>2</sub>-EA metabolite peaks at  $m/z = 418$ . Quantitation of COX-2 activity is accomplished by comparing the area of the PGH<sub>2</sub>-EA metabolite peak to that of the internal standard.

## REFERENCES

All references, U.S. Patents, non U.S. Patents, journal articles, and newspaper articles referred to herein are hereby made part of the specification of the present patent and incorporated herein in their entirety by reference. This includes, but is not limited to the following references:

Bisogno, T., Melck, D., De Petrocellis, L., & Di Marzo, V. (1999) *J. Neurochem.* 72:2113-2119.

DuBois, RN, Radhika, A, Reddy, BS, Entingh, AJ. (1996) *Gastroenterology* 110:1259.

Cohen, et al. U.S. Patent No. 5,589,575, *Purification of hapten-carrier generated antibodies*.

Cayman Chemical, 1180 E. Ellsworth Road, Ann Arbor, MI 48108; 800-364-9897; *Prostaglandin E<sub>2</sub> affinity purification kit*, Catalog No. 514018.

Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin D, Mandelbaum A, Etinger A, & Mechoulam R (1992) *Science* 258:1946-1949.

- Eberhart, CE, Coffey, RJ, Radhika, A, *et al.* (1994) *Gastroenterology* 107:1183.
- Fosslien, E. (2000) *Ann. Clin. Lab. Sci.* 30:3-21.
- Jeon, YJ, Yang, KH, Pulaski, JT, & Kaminski, NE. (1996) *Mol. Pharmacol.* 50:334-341.
- 5 Kalgutkar, AS, Kozak, KR, Crews, BC, Hochgesang, Jr. GP, & Marnett, LJ. (1998) *J. Med. Chem.* 41: 4800-4818.
- Kalgutkar, AS, Crews, BC, Rowlinson, SW, Marnett, AB, Kozak, KR, Remmel, RP, & Marnett, LJ. (2000) *Proc. Natl. Acad. Sci. USA* 97: 925-930.
- Kennedy, BP, Chan, C-C, Culp, SA, & Cromlish, WA. (1993) *Biochem. Biophys. Res. Commun.* 197:494-500.
- Kozak, KR, Rowlinson, SW, & Marnett, LJ. (2000) *J. Biol. Chem.* 275:33744-33749.
- Landino, LM, Crews, BC, Timmons, MD, Morrow, JD, & Marnett, LJ. (1996) *Proc. Natl. Acad. Sci. USA* 93:15069-15074.
- MacPherson, JC, Pavlovich, JG, & Jacobs, RS. (1996) *Biochim. Biophys. Acta* 1303:127-136.
- 15 Marnett, LJ, Siedlik, PH, Ochs, RC, Pagels, WD, Das, M, Honn, KV., Warnock, RH, Tainer, BE, & Eling, TE. (1984) *Mol. Pharmacol.* 26:328-335.
- Masferrer, JL, Zweifel, BS, Manning, PT, Hauser, SD, Leahy, KM., Smith, WG, Isakson, PC, & Seibert, K. (1994) *Proc. Natl. Acad. Sci. USA* 91, :28-3232.
- 20 McCafferty, J., Hoogenboom, H., & Chiswell, D. (1996) *Antibody Engineering, a Practical Approach*, IRL Press @ Oxford University Press.
- Mernaugh, R. & Mernaugh, G. (1994) *Methods for the Production of Monoclonal Antibodies*, in **Molecular Methods in Plant Pathology**, Ed by RP Singh and US Singh, pg. 343-365.

Odenwaller, R, Chen, Y-NP., & Marnett, LJ. (1990) *Methods. Enzymol.* 187: 479-485.

Rowlinson, SW, Crews, BC., Lanzo, CA, & Marnett, LJ. (1999) *J. Biol. Chem.* 274: 23305-23310.

5 Rowlinson, SW, Crews, BC, Goodwin, DC, Schneider, C, Gierse, JK, & Marnett, L. J. (2000) *J. Biol. Chem.* 274:6586-6591

Sheehan, KM, Sheahan, K, O'Donoghue, DP, *et al.* (1999) *JAMA* 282:1254.

Sheng, H, Shao, J, Kirkland, SC, *et al.* (1997) *J. Clin. Invest.* 99:2254.

So, O-Y, Scarafia, LE, Mak, AY, Callan, OH, & Swinney, DC (1998) *J. Biol. Chem.* 273:5801-5807.

Stella, N., Schweitzer, P., & Piomelli, D. (1997) *Nature* 388:773-778.

Tsujii, M, Kawano, S, Du Bois, RN. (1997) *Proc. Natl. Acad. Sci. USA* 94:3336.

Vane, JR, Mitchell, JA, Appleton, I, Tomlinson, A, Bishop-Bailey, D, Croxtall, J, & Willoughby, DA (1994) *Proc. Natl. Acad. Sci. USA* 91:2046-2050.

15 Wadleigh, DJ, Reddy, ST, Kopp, E, Ghosh, S, & Herschman, HR (2000) *J. Biol. Chem.* 275:6259-6266.

Yu, M, Ives, D, & Ramesha CS (1997) *J. Biol. Chem.* 272:21181-21186.

20 This invention thus being described, it will be obvious that the same may be varied in many ways. Such variations are not to be regarded as a departure from the spirit and scope of the present invention, and all such modifications as would be obvious to one of ordinary skill in the art are intended to be included within the scope of the claims following the detailed description of the invention. The present invention is not bound by theory or mechanism. Thus, although there have been

Inventors: Marnett and Kozak  
Attorney Docket No.: N-7362-RSM  
Customer No.: 23456

described particular embodiments of the present invention of a new and useful "Method for *in vitro* and *in vivo* determination of COX-2 activity", it is not intended that such references be construed as limitations upon the scope of this invention except as set forth in the following claims.